# Organic acids promote phosphorus release from Mollisols with different organic matter contents

Xiaoyan Yang<sup>1</sup>, Chuandong Zhang<sup>2</sup>, Haiping Gu<sup>1</sup>, Xiangwei Chen<sup>3</sup>, Erhui Guo<sup>1</sup>\*

**Citation:** Yang X., Zhang C., Gu H., Chen X., Guo E. (2021): Organic acids promote phosphorus release from Mollisols with different organic matter contents. Soil & Water Res., 16: 59–66.

**Abstract:** Organic acids could improve the phosphorus (P) availability through enhancing the release of inorganic phosphorus ( $P_i$ ) in the soil. However, the effects of organic acids on the  $P_i$  release are still poorly understood, especially from soils with different organic matter contents. Here, a biochemically produced humic acid and P fertiliser were added to the soil to modify the content of the soil organic matter (SOM) and soil P, respectively. And then the soil samples were incubated at 25 °C for 30 days. The release of  $P_i$  fractions (such as  $H_2O-P_i$ ,  $NaHCO_3-P_i$ ,  $NaOH-P_i$ ,  $HCl-P_i$ , and Residual-P) from the soils with different organic matter contents in the presence of citric, oxalic, and malic acids was evaluated using a sequential chemical fractionation method. The results showed that the release of the  $NaHCO_3-P_i$ ,  $NaOH-P_i$ , and  $HCl-P_i$  fractions also showed a decreasing trend with an increasing content of soil organic matter, and more  $NaOH-P_i$  than the other  $P_i$  fractions was generally released in the presence of organic acids. Considering the types of organic acids, oxalic acid and malic acid most effectively and least effectively released  $P_i$ , respectively. The path analysis indicated that the  $NaOH-P_i$  release had the highest direct and indirect effects on the total inorganic P ( $TP_i$ ) release.  $NaOH-P_i$  was, therefore, the most effective source of  $P_i$  in the Mollisols.

Keywords: available phosphorus; black soil; correlation analysis; path analysis; phosphorus fractions

Phosphorus (P) is an essential nutrient required by crops. P deficiencies are relatively more common in the early part of the growing season (Hopkins et al. 2008), and, therefore, can limit crop growth in a wide range of soils across the world (Holford 1997). Phosphate fertilisers are applied to soils to maintain and improve crop yields (Romanyà et al. 2017). However, excessive P fertiliser applications cause P to accumulate in the soil (Ahmed et al. 2019) and cause various other environmental problems (Daniel et al. 1998). P is present in the soil in various forms that have a different availability and solubil-

ity. A sequential P fractionation scheme based on a method developed by Hedley et al. (1982) has been used in many studies of soil P fractions (Rakotoson et al. 2015; Yan et al. 2017; Yang et al. 2019a). This method allows one to determine the P availability in soils associated with vegetation (DeBruler et al. 2019; Liao et al. 2020), management regimes (Shi et al. 2015), and land use change (Sun et al. 2019). Among them, inorganic P ( $P_i$ ) in the soil is the preferred source of P for plants and may pose greater risks for eutrophication than organic P (Zhang et al. 2018), because the potential bioavailability of

<sup>&</sup>lt;sup>1</sup>School of Forestry, Henan Agricultural University, Zhengzhou, P.R. China

<sup>&</sup>lt;sup>2</sup>The Third Geological Exploration Institute, Henan Bureau of Geo-Exploration Mineral Development, Zhengzhou, P.R. China

 $<sup>^3</sup>$ School of Forestry, Northeast Forestry University, Harbin, P.R. China

<sup>\*</sup>Corresponding author: guoerhui@126.com

This work was supported by National Natural Science Foundation of China (No. 41701360).

organic phosphorus in the various fractions lacks experimental validation (Turner et al. 2005). So, it is important to study soil P<sub>i</sub> fractions to improve our understanding of soil P dynamics.

Organic acids exuded by plants and soil microbiota (Romanyà et al. 2017) can transform insoluble P into plant-available forms and, therefore, affect the P bioavailability (Wang et al. 2018; Yang et al. 2019a). However, the abilities of organic acids to solubilise P are dependent on the soil type, the soil organic matter (SOM) content, and the structures and concentrations of the organic acids themselves (Oral & Uygur 2018). SOM is one of the important constituents in the soil in terms of maintaining the solubility of P in the soil (Yan et al. 2016; Romanyà et al. 2017). The effects of SOM on the soil P availability have been widely reported with contradictory results (Guan et al. 2006; Hua et al. 2008; Clarholm et al. 2015). In addition, the release of organic acids destabilises the organic matter in organomineral complexes and causes decomposition that affects the effectiveness of the SOM onto the P bioavailability (Clarholm et al. 2015). It is, therefore, important to understand the effects of the interactions between the organic acids and the SOM on the availability of P to ensure that phosphate fertilisers are used effectively. Few studies on this have been performed.

Therefore, the main aim was to identify the roles organic acids play in releasing P from different  $P_i$  fractions in Mollisols with different SOM contents which were prepared by adding humic acids to the natural soil. The contribution of the different  $P_i$  fractions to the release of the total  $P_i$  ( $TP_i$ ) was assessed by performing a path analysis.

We hypothesised that different types of organic acids, due to the different affinities for the dominant bridge-forming polyvalent metal cations in the SOM, as well as the suitable size and sufficient lipophilicity to penetrate hydrophobic SOM structures (Piccolo 2002), promoted different levels of P release in soils with different SOM contents. In addition, the effect of the individual  $P_i$  fraction on the release of the total  $P_i$  varied, and this variation, to some extent, depended on its content and bioavailability.

#### MATERIAL AND METHODS

# Soil sampling

The experiments were performed using soil from Keshan Farm, which is in a typical black soil area in northeast China (N 48°12′–48°23′, E 125°08′–125°37′).

The soil was classified as a Mollisol. Soil samples from 40-45 cm deep were collected from five randomly assigned points on the arable land used for a seasonal maize crop to obtain a lower SOM content for the simulation experiment. The soil samples were dried and then passed through a 2 mm sieve before being used. The bulk density was measured using the core sampling method, using a core size of 100 cm<sup>3</sup> (Chen 2005). The soil pH was measured using a pH meter with a glass electrode in a 1:2.5 (w/v) soil-to-deionised water ratio (Chen 2005). The soil organic carbon content was determined using a Multi EA 400 elemental analyser (Vario EL cube, Elementar, Germany), and a conversion factor of 1.724 was used to calculate the SOM (Chen 2005). The soil sample was digested in a mixture of H<sub>2</sub>SO<sub>4</sub> and HClO<sub>4</sub> (Parkinson & Allen 1975) to obtain the total P fraction. The available P was extracted with 0.03 mol/L NH<sub>4</sub>F-0.025 mol/L HCl and determined using the colorimetric method developed by Murphy and Riley (1962). The soil bulk density was 1.17 g/cm<sup>3</sup>, the pH was 6.18, the SOM content was 20.5 g/kg, the total P content was 442 mg per kg, and the available P content was 5.04 mg/kg.

# **Experimental design**

Preparation of "spare" soil samples with different SOM contents. Biochemically produced humic acids purchased from Aladdin Biochemical Technology Co. (Shanghai, China) were added to the soil to modify the SOM content. The humic acids have a negligible total P content and a C content of 36.7%. Humic acids of 0%, 4%, 8%, 11% (humic acid/ soil sample (w/w)) were added to 300 g of soil to give samples with SOM contents of 20.5, 44.6, 68.9, and 99.6 g/kg, which were labelled SOM<sub>1</sub>, SOM<sub>2</sub>, SOM<sub>3</sub>, and SOM<sub>4</sub>, respectively. A KH<sub>2</sub>PO<sub>4</sub> fertiliser at a rate of 200 mg P/kg of soil was then added to each soil to increase the P content (Jalali & Ranjbar 2010). Deionised water was added to adjust the water content to 70% of the field capacity and then replenished every other day using the gravimetric method during incubation. The samples were incubated at 25 °C for 30 days, because the contents of the P fractions in the soil samples treated with the KH<sub>2</sub>PO<sub>4</sub> and humic acids were almost stable after being incubated for 30 days in the pre-experiment. Each treatment was undertaken in triplicate, thus, twelve soil samples were incubated. After incubation, the three replicate soils of the same treatment were thoroughly mixed, dried, and then were passed through a 2 mm sieve before use. These samples are called "spare" soil samples below, and

there are four "spare" soil samples (namely,  $SOM_1$ ,  $SOM_2$ ,  $SOM_3$ , and  $SOM_4$ ) in total.

"Spare" soil analysis. A sequential P fraction extraction was performed using a modified version of a fractionation scheme developed by Hedley et al. (1982). Four aliquots of each soil sample were subjected to the extraction procedure. A 0.5 g aliquot of a soil sample was placed in a 50 mL centrifuge tube and sequentially extracted with 30 mL of  $H_2O$ , 0.5 mol/L of NaHCO<sub>3</sub> (at pH 8.5), 0.1 mol/L of NaOH, and then 1 mol/L of HCl, for 16 h with each extractant. The soil residue was then digested in a mixture of H2SO4 and HClO4 to obtain the P content of the solid residue (later labelled Residual-P) to be determined. The P concentrations in all the above-mentioned extracts were determined using the colorimetric method developed by Murphy and Riley (1962).

The soil pH, and the contents of the total P, available P, and SOM in the "spare" soil were determined as described above (see the part of the soil sampling).

Organic acid amendments. A 1.0 g aliquot of "spare" soil was extracted for 30 min with 20 mL of a solution containing oxalic acid, citric acid, or malic acid at a concentration of 2.5 mmol/L, respectively. These are the most important organic acids exuded by plants and the mycorrhizae (Clarholm et al. 2015). The organic acid concentration was selected taking the concentrations found in rhizosphere samples by Santos et al. (2017) into consideration. The soil suspension was centrifuged at 5000 rpm for 10 min and passed through a 0.45 µm membrane filter. The total inorganic P (TP<sub>i</sub>) concentration in the supernatant was determined using the molybdenum blue colorimetric method (Murphy & Riley 1962). The amount of TP<sub>i</sub> release was defined as the difference between the amount of TP<sub>i</sub> extracted with and without the organic acids in the extractant solution (Yang et al. 2019a). Each soil sample that had been extracted with an organic acid was then washed with alcohol to remove the residual organic acid between the soil particles (Liu et al. 2009), then the P<sub>i</sub> fractions were determined using the sequential extraction method modified by Hedley et al. (1982). This divided the P<sub>i</sub> into the H<sub>2</sub>O-P<sub>i</sub>, NaHCO<sub>3</sub>-P<sub>i</sub>, NaOH-P<sub>i</sub>, HCl-P<sub>i</sub>, and Residual-P fractions. The amount of each P<sub>i</sub> fraction release was defined as the difference between the P<sub>i</sub> fraction concentration found when the soil had and had not been treated with the organic acids (Yang et al. 2019a). Each treatment was performed on four separate soil aliquots.

### Statistical analyses

A one-way analysis of variance (ANOVA) was performed and the mean values were compared using t-tests (P < 0.05). Simple correlation analyses were performed to investigate the interactions between the release of  $TP_i$  and the release of the different  $P_i$  fractions. Path analyses were performed to attempt to explain the contributions of the release of the different  $P_i$  fractions from the  $TP_i$ . All the statistical analyses were performed using SPSS 16.0 (IBM, Armonk, NY, USA), and a significance threshold of P < 0.05 was used. The figures were drawn using SigmaPlot 10.0 (Systat Software, San Jose, CA, USA).

# **RESULTS**

**Properties of P in different soils.** No significant differences were found between the TP contents within the soil samples with the different SOM contents (P < 0.05), indicating that the addition of humic acid did not affect the soil TP contents (Table 1). This indicated that the humic acid added to the soil samples contained almost no P. The available P contents increased as the SOM content increased.

Most of the  $P_i$  fractions were in the moderately labile NaOH- $P_i$  and the most stable Residual-P frac-

Table 1. Soil pH and P contents in the soils with different organic matter contents

Soil samples	pH -	TP	AP	P <sub>i</sub> fractions (mg/kg)						
		(mg/kg)		H <sub>2</sub> O-P <sub>i</sub>	NaHCO <sub>3</sub> -P <sub>i</sub>	NaOH-P <sub>i</sub>	HCl-P <sub>i</sub>	Residual-P		
SOM <sub>1</sub>	6.14	616ª	81.32 <sup>c</sup>	41.84ª	76.08 <sup>a</sup>	121.76 <sup>c</sup>	42.67 <sup>b</sup>	134.21ª		
$SOM_2$	5.80	621 <sup>a</sup>	84.77 <sup>bc</sup>	$39.30^{b}$	73.81 <sup>a</sup>	$129.25^{bc}$	$43.75^{b}$	131.29 <sup>a</sup>		
$SOM_3$	5.58	623 <sup>a</sup>	86.83 <sup>ab</sup>	$41.76^{a}$	$68.47^{\rm b}$	$133.00^{b}$	$44.10^{b}$	129.45 <sup>a</sup>		
$SOM_4$	5.19	620 <sup>a</sup>	90.81 <sup>a</sup>	$42.48^{a}$	53.61 <sup>c</sup>	156.83 <sup>a</sup>	47.18 <sup>a</sup>	123.56 <sup>a</sup>		

The different lowercase letters indicate a significance among the different soil organic matter (SOM) content soils at P < 0.05; TP – total phosphorus; AP – available phosphorus; SOM $_1$  – 20.5 g/kg; SOM $_2$  – 44.6 g/kg; SOM $_3$  – 68.9 g/kg; SOM $_4$  – 99.6 g/kg;  $P_i$  – inorganic phosphorus

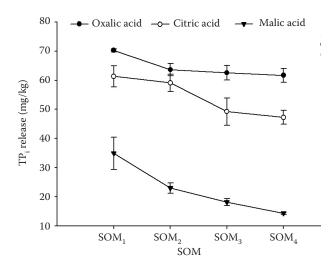


Figure 1. The amounts of total inorganic P  $(TP_i)$  release from the different organic matter content soils  $SOM_1, SOM_2, SOM_3$  and  $SOM_4$  represent 20.5, 44.6, 68.9, and 99.6 g/kg, respectively

tion. As the SOM content increased, the NaHCO $_3$ -P $_i$  content decreased gradually and it was higher for SOM $_1$  than for SOM $_2$ -SOM $_4$ . In contrast to the NaHCO $_3$ -P $_i$  content, the NaOH-P $_i$  and HCl-P $_i$  content increased as the SOM content increased, while the H $_2$ O-P $_i$  and Residual-P content fluctuated as the SOM content varied.

 $TP_i$  release. The organic acids could solubilise the  $TP_i$  significantly (Figure 1). When the SOM content was the same, more  $TP_i$  was released from all four

soil samples by the oxalic acid compared to the other acids. Citric acid released the second largest amount of  $TP_i$  and malic acid released the least. Significant differences were found between the amounts of  $TP_i$  released by the three organic acids from all the soil samples (P < 0.05). This indicated that citric acid was more effective in releasing the  $TP_i$  than both the citric and malic acids.

By the same organic acid, the amount of  $TP_i$  release decreased significantly as the SOM content increased (P < 0.05). The degree to which the amount of the  $TP_i$  release decreased as the SOM content increased was lowest for the oxalic acid, the next lowest was for the citric acid, and highest for the malic acid. This revealed that the ability of malic acid to release the  $TP_i$  was more affected by the organic matter content than that of the oxalic and citric acid.

Release of different  $P_i$  fractions. Table 2 shows that more NaOH- $P_i$  than the other fractions was released except for with malic acid, which enhanced the release of NaHCO<sub>3</sub>- $P_i$  more than the other fractions, followed by NaHCO<sub>3</sub>- $P_i$ , then HCl- $P_i$ , and finally Residual- $P_i$ .

At the same SOM content, the amount of the  $H_2O-P_i$  release had a negative value, indicating that the  $H_2O-P_i$  content was higher in the presence of organic acids. In contrast, the amounts of the NaHCO<sub>3</sub>-P<sub>i</sub>, NaOH-P<sub>i</sub>, and HCl-P<sub>i</sub> released by the organic acids had positive values, and were the highest for oxalic acid, the next highest for citric acid, and the lowest for malic

Table 2. The amounts of the inorganic phosphorus (P<sub>i</sub>) fractions released by the organic acids (in mg/kg)

Soil samples	Organic acids types	H <sub>2</sub> O-P <sub>i</sub>	NaHCO <sub>3</sub> -P <sub>i</sub>	NaOH-P <sub>i</sub>	HCl-P <sub>i</sub>	Residual-P
	oxalic acid	-17.61 <sup>aB</sup>	39.15 <sup>aA</sup>	72.96 <sup>aA</sup>	10.10 <sup>aA</sup>	5.67 <sup>aA</sup>
$SOM_1$	citric acid	$-12.55^{bC}$	$29.23^{\mathrm{bA}}$	$60.59^{bA}$	$8.44^{\mathrm{bA}}$	5.58 <sup>aA</sup>
	malic acid	$-6.10^{\rm cC}$	$21.36^{\mathrm{cA}}$	$18.33^{cA}$	$2.55^{\mathrm{cA}}$	$4.92^{aA}$
$SOM_2$	oxalic acid	$-23.468^{aA}$	$28.18^{aB}$	68.96 <sup>aAC</sup>	$9.41^{\mathrm{aAB}}$	$4.34^{aA}$
	citric acid	$-18.68^{bB}$	$24.26^{\mathrm{bB}}$	$57.18^{bAB}$	$8.39^{\mathrm{bA}}$	$4.03^{aA}$
	malic acid	$-8.48^{cB}$	$13.70^{cB}$	$13.70^{\mathrm{cB}}$	$2.66^{\mathrm{cA}}$	$3.11^{bA}$
$SOM_3$	oxalic acid	$-24.38^{aA}$	$20.38^{aC}$	67.66 <sup>aBC</sup>	$9.01^{aB}$	$3.64^{aA}$
	citric acid	$-23.95^{aA}$	17.25 <sup>bC</sup>	$56.41^{\text{bAB}}$	$7.80^{aA}$	$3.08^{aA}$
	malic acid	$-9.23^{\mathrm{bB}}$	$13.33^{cB}$	$13.29^{cB}$	$2.24^{\mathrm{bA}}$	$3.00^{aA}$
$SOM_4$	oxalic acid	$-26.53^{aA}$	$18.74^{\mathrm{aC}}$	$64.00^{aB}$	$8.79^{aB}$	$3.08^{aA}$
	citric acid	$-23.33^{aA}$	$17.11^{bC}$	$52.53^{bB}$	$7.89^{aA}$	$3.36^{aA}$
	malic acid	$-12.93^{bA}$	$13.50^{\mathrm{cB}}$	$9.71^{\mathrm{cB}}$	$2.58^{bA}$	$2.05^{aA}$

The different lowercase letters indicate a significance among the different organic acids types at P < 0.05; the different capital letters indicate a significance among the different soils of the soil organic matter (SOM) content at P < 0.05; SOM<sub>1</sub> – 20.5 g/kg; SOM<sub>2</sub> – 44.6 g/kg; SOM<sub>3</sub> – 68.9 g/kg; SOM<sub>4</sub> – 99.6 g/kg

Table 3. Correlation matrices between the release of the total inorganic P (TP<sub>i</sub>) and the inorganic phosphorus (P<sub>i</sub>) fractions

	$TP_i$	H <sub>2</sub> O-P <sub>i</sub>	NaHCO <sub>3</sub> -P <sub>i</sub>	NaOH-P <sub>i</sub>	HCl-P <sub>i</sub>	Residual-P
$TP_i$	1.000	-0.649**	0.747**	0.958**	0.917**	0.302*
$H_2O-P_i$		1.000	-0.147	-0.751**	-0.741**	0.052
NaHCO <sub>3</sub> -P <sub>i</sub>			1.000	0.664**	0.629**	0.427
NaOH-P <sub>i</sub>				1.000	0.957**	0.218
HCl-P <sub>i</sub>					1.000	0.166
Residual-P						1.000

<sup>\*, \*\*</sup>Significant at *P* < 0.05, 0.01

Table 4. Direct effects and indirect effects of the individual inorganic phosphorus  $(P_i)$  release on the total inorganic  $P(TP_i)$  release

	Direct-path	Indirect-path coefficient					
	coefficient	$H_2O$ - $P_i$	→NaHCO <sub>3</sub> -P <sub>i</sub>	→NaOH-P <sub>i</sub>	$\rightarrow$ HCl-P <sub>i</sub>	→Residual-P	
H <sub>2</sub> O-P <sub>i</sub>	-0.002		-0.026	-0.590	-0.033	0.002	
NaHCO <sub>3</sub> -P <sub>i</sub>	0.176*	0.000		0.522	0.028	0.020	
NaOH-P <sub>i</sub>	0.786**	0.002	0.117		0.043	0.010	
HCl-P <sub>i</sub>	0.045	0.002	0.111	0.752		0.008	
Residual-P	0.048	0.000	0.075	0.171	0.007		

<sup>\*, \*\*</sup>Significant at *P* < 0.05. 0.01

acid. However, the amounts of the Residual-P release were not significantly different for the soil samples.

For the same organic acid, the absolute amount of the  $H_2O-P_i$  release increased, while the amounts of the  $NaHCO_3-P_i$ ,  $NaOH-P_i$ , and  $HCl-P_i$  release decreased as the SOM content increased (but with different levels of significance). The amounts of the Residual-P release were not significantly different for the soil samples with the different SOM contents (Table 2).

**Correlation analysis.** The release of  $TP_i$  positively correlated with that of the  $P_i$  fractions (P < 0.05) (Table 3), and the NaOH- $P_i$  release made the strongest correlation. For different  $P_i$  fractions, significant correlations were found between the release of the NaHCO<sub>3</sub>- $P_i$ , NaOH- $P_i$ , and HCl- $P_i$ . The release of the  $H_2O-P_i$  was significantly negatively correlated with that of the NaOH- $P_i$  and HCl- $P_i$  (P < 0.01).

**Path analysis.** A path analysis was performed to assess the direct and indirect effects of the  $P_i$  fractions release from the  $TP_i$  (Table 4). The direct-path coefficients indicated that the release of the NaHCO<sub>3</sub>- $P_i$  and NaOH- $P_i$  directly and significantly affected the  $TP_i$  release (P < 0.05) and that the NaOH- $P_i$  release made a larger contribution to the  $TP_i$  release (P < 0.01) than did the NaHCO<sub>3</sub>- $P_i$  release. The indirect-path coefficients indicated that the indirect

effects of the  $P_i$  fractions release on the  $TP_i$  release were stronger for the release of the NaOH- $P_i$  than that of the other fractions. The release of the  $H_2O-P_i$ ,  $HCl-P_i$ , and Residual-P made small contributions to the  $TP_i$  release.

#### **DISCUSSION**

In our study, the abilities of organic acids to release the Pi decreased gradually as the SOM content increased. In a previous study of P adsorption in Mollisols, Yang et al. (2019b) found that the maximum P adsorption capacity and soil P adsorption index increased with the SOM content. This ultimately caused smaller amounts of Pi to be released by the organic acids as the SOM content increased. Gerke and Hermann (1992) and Oburger et al. (2011) found that organic-inorganic complexes formed between the humus (part of the SOM) and the iron/aluminium species provided important sites for the adsorption of P<sub>i</sub>, giving, for example, ternary humic-Al (Fe)-P<sub>i</sub> complexes. This would increase the amount of P that would be adsorbed and markedly decrease the P<sub>i</sub> concentrations in the soil solution. However, Summerhays et al. (2015) found that adding organic acids created conditions similar to the conditions found

in the presence of SOM, minimising the P solubilisation from the soil cations. This was also found for alkaline/calcareous soils in which excess cations precipitate or adsorb labile P. In conclusion, the data suggested that the high organic acid contents of a soil with a high SOM content did not improve the P<sub>i</sub> bioavailability to the extent offered by the high organic acid contents of the soil with a relatively low SOM content. Exogenous organic acids appeared to most effectively improve the release of P from the soil with a low to moderate SOM content. Kpomblekou et al. (2003) found that acids were more effective in releasing P from low and medium reactive phosphate rocks than from high reactive phosphate rocks. We used soils with high P contents through the addition of external P. Future studies should be performed using a soil with a low P content.

For the inorganic P fractions, the content of the NaOH-P<sub>i</sub> in the "spare" soil samples and the amounts of the NaOH-P<sub>i</sub> release in the presence of the organic acids were higher than those of the other P<sub>i</sub> fractions, indicating that the NaOH-P<sub>i</sub> fraction could be a source of P<sub>i</sub> to the plants. The path analysis results indicated that release of the NaOH-P<sub>i</sub> fraction had the strongest direct and indirect effects on the TP<sub>i</sub> release. The simple correlation analysis results agreed with that result, the highest correlation coefficient being for the relationship between the release of the NaOH-P<sub>i</sub> and the TPi. This confirmed that the NaOH-Pi was a more effective source than the other fractions in the Mollisol at an intermediate time-scale. The strong direct effect of the NaHCO3-Pi release on the TPi release and the correlation coefficient between them indicated that the NaHCO3-Pi could make a strong contribution to the plant-available P in agricultural soils in the short term because it was rapidly desorbed from the soil particle surfaces (Cross & Schlesinger 1995). Verma et al. (2005) found that the contributions of the NaHCO<sub>3</sub>-P<sub>i</sub> to the P availability to the plant uptake of P were higher for acidic soils that had been continually fertilised with mineral and organic fertilisers than for other soils. Yang et al. (2006) also found that the NaHCO<sub>3</sub>-P<sub>i</sub> and NaOH-P<sub>i</sub> contributed more than the other P fractions to the available P (Bray P) in red soil regions. The lower direct-path and indirect-path coefficients for the release of the H<sub>2</sub>O-P<sub>i</sub>, HCl-P<sub>i</sub> and Residual-P could probably be explained by the lower contents of the H<sub>2</sub>O-P<sub>i</sub> and HCl-P<sub>i</sub>, and the smaller release of the HCl-P<sub>i</sub> and Residual-P by the organic acids. These results were consistent with the moderately resistant HCl- $P_i$  fraction and the highly resistant Residual-P fraction being less effective than the NaHCO<sub>3</sub>- $P_i$  and NaOH- $P_i$  as sources of  $P_i$  to plants (Rose et al. 2010). In addition, a significant negative correlation was found between the release of the  $H_2O$ - $P_i$  and NaOH- $P_i$ . This explained the discovery by Tiessen et al. (1984) that most labile P originated in the NaOH- $P_i$  fraction.

In our study, the high doses of humic acids, as well as P fertilisers, were chosen to explore the soil P release with different organic matter contents. The significant results are then obvious and these results have important theoretical implications for our understanding of the P release mechanism. Despite this, considering the lower amounts of P fertilisers and organic matter in agricultural practice, further work is needed to determine the effect of low doses of humic acids as well as P fertilisers under different gradients on the soil P release.

#### **CONCLUSION**

The moderately labile NaOH-Pi and stable Residual-P fraction contents were higher than the other P<sub>i</sub> fraction contents of Mollisols with different SOM contents. The effects of the organic acids on the P<sub>i</sub> release were related to the SOM contents. Oxalic, citric, and malic acids increased the release of the TP<sub>i</sub> and the individual P<sub>i</sub> fractions, and the abilities of the acids to release P decreased as the SOM content increased. In other words, the higher the SOM content, the weaker the abilities of the organic acids to release the different P<sub>i</sub> fractions. For the P<sub>i</sub> fractions, more NaOH-P<sub>i</sub> than the other P<sub>i</sub> fractions was released by the organic acids. The amounts of the NaHCO<sub>3</sub>-P<sub>i</sub> release were the second highest. The path analysis results indicated that the organic acids increased the availability of P mainly by releasing NaOH-Pi and NaHCO3-Pi from the Mollisols.

**Acknowledgement:** We thank G. Thomas, Ph.D., from Liwen Bianji, Edanz Group China (www.liwenbianji.cn/ac), for editing the English text of a draft of this manuscript.

# **REFERENCES**

Ahmed W., Jing H., Kaillou L., Qaswar M., Zhang H. (2019): Changes in phosphorus fractions associated with soil chemical properties under long-term organic and inorganic fertilization in paddy soils of southern China. PLoS ONE, 14: e0216881.

- Chen L.X. (2005): Soil Experiment and Practice Tutorials. Harbin, Northeast Forestry University Press.
- Clarholm M., Skyllberg U., Rosling A. (2015): Organic acid induced release of nutrients from metal-stabilized soil organic matter The unbutton model. Soil Biology and Biochemistry, 84: 168–176.
- Cross A.F., Schlesinger W.H. (1995): A literature review and evaluation of the Hedley fractionation: Applications to the biogeochemical cycle of soil phosphorus in natural ecosystems. Geoderma, 64: 197–214.
- Daniel T.C., Sharpley T.C., Lemunyon J.L. (1998): Agricultural phosphorus and eutrophication: A symposium overview. Journal of Environmental Quality, 27: 251–257.
- DeBruler D.G., Schoenholtz S.H., Slesak R.A., Strahm B.D., Harrington T.B. (2019): Soil phosphorus fractions vary with harvest intensity and vegetation control at two contrasting Douglas-fir sites in the Pacific northwest. Geoderma, 350: 73–83.
- Gerke J., Hermann R. (1992): Adsorption of orthophosphate to humic-Fe-complexes and to amorphous Fe-oxide. Journal of Soil Science and Plant Nutrition, 155: 233–236.
- Guan X.H., Shang C., Chen G.H. (2006): Competitive adsorption of organic matter with phosphate on aluminum hydroxide. Journal of Colloid and Interface Science, 296: 51–58.
- Hedley M.J., Stewart J.W.B., Chauhan B.S. (1982): Changes in inorganic and organic soil phosphorus fractions induced by cultivation practices and by laboratory incubations. Soil Science Society of America Journal, 46: 970–976.
- Holford I.C.R. (1997): Soil phosphorus: Its measurement, and its uptake by plants. Australian Journal of Soil Research, 35: 227–239.
- Hopkins B.G., Rosen C.J., Shiffler A.K., Taysom T.W. (2008): Enhanced efficiency fertilizers for improved nutrient management: potato (*Solanum tuberosum*). Crop Management, 7: 1–16.
- Hua Q.X., Li J.Y., Zhou J.M., Wang H.Y., Du C.W., Chen X.Q. (2008): Enhancement of phosphorus solubility by humic substances in Ferrosols. Pedosphere, 18: 533–538.
- Jalali M., Ranjbar F. (2010): Aging effects on phosphorus transformation rate and fractionation in some calcareous soils. Geoderma, 155: 101–106.
- Kpomblekou A.K., Tabatabai M.A. (2003): Effect of low-molecular weight organic acids on phosphorus release and phytoavailabilty of phosphorus in phosphate rocks added to soils. Agriculture, Ecosystems and Environment, 100: 275–284.
- Liao D., Zhang C., Li H., Lambers H., Zhang F. (2020): Changes in soil phosphorus fractions following sole cropped and intercropped maize and faba bean grown on calcareous soil. Plant and Soil, 448: 587–601.

- Liu L., Liang C.H., Wang Q., Du L.Y., Wu Y.M., Han W. (2009): Effects of low-molecular-weight organic acids on soil phosphorus release. Plant Nutrition and Fertility Science, 15: 593–600.
- Murphy J., Riley J.P. (1962): A modified single solution method for the determination of phosphate in natural waters. Analytica Chimica Acta, 27: 31–36.
- Oburger E., Jones D.L., Wenzel W.W. (2011): Phosphorus saturation and pH differentially regulate the efficiency of organic acid anion-mediated P solubilization mechanisms in soil. Plant Soil, 341: 363–382.
- Oral A., Uygur V. (2018): Effects of low-molecular-mass organic acids on P nutrition and some plant properties of *Hordeum vulgare*. Journal of Plant Nutrition, 41: 1482–1490.
- Parkinson J.A., Allen S.E. (1975): A wet oxidation procedure suitable for determination of nitrogen and mineral nutrients in biological material. Communications in Soil Science and Plant Analysis, 6: 1–11.
- Piccolo A. (2002): The supramolecule structure of humic substances. A novel understanding of humic substances and implications for soil science. Advances in Agronomy, 57: 57–134.
- Rakotoson T., Six L., Razafimanantsoa M.P., Rabeharisoa L., Smolders E. (2015): Effects of organic matter addition on phosphorus availability to flooded and nonflooded rice in a P-deficient tropical soil: a greenhouse study. Soil Use and Management, 31: 10–18.
- Romanyà J., Blanco-Moreno J.M., Sans F.X. (2017): Phosphorus mobilization in low-P arable soils may involve soil organic C depletion. Soil Biology and Biochemistry, 113: 250–259.
- Rose T.J., Hardiputra B., Rengel Z. (2010): Wheat, canola and grain legume access to soil phosphorus fractions differs in soils with contrasting phosphorus dynamics. Plant Soil, 326: 159–170.
- Santos S.R., Silva E.D.B., Alleoni L.R.F., Grazziotti P.H. (2017): Citric acid influence on soil phosphorus availability. Journal of Plant Nutrition, 40: 2138–2145.
- Shi Y.C., Ziadi N., Messiga A.J., Lalande R., Hu Z.Y. (2015): Soil phosphorus fractions change in winter in a cornsoybean rotation with tillage and phosphorus fertilization. Pedosphere, 25: 1–11.
- Summerhays J.S., Hopkins B.G., Jolley V.D., Hill M.W., Ransom C.J., Brown T.R. (2015): Enhanced phosphorus fertilizer (Carbond P) supplied to maize in moderate and high organic matter soils. Journal of Plant Nutrition, 38: 1359–1371.
- Sun X., Li M., Wang G., Drosos M., Liu F., Hu Z. (2019): Response of phosphorus fractions to land-use change followed by long-term fertilization in a sub-alpine humid

- soil of Qinghai–Tibet plateau. Journal of Soils and Sediments, 19: 1109–1119.
- Tiessen H., Stewart J.W.B., Cole C.V. (1984): Pathways of phosphorus transformation in soils of differing pedogenesis. Soil Science Society of America Journal, 48: 853–858.
- Turner B.L., Cademenun B.J., Condron L., Newman S. (2005): Extraction of soil organic phosphorus. Talanta, 66: 294–306.
- Verma S., Subehia S.K., Sharma S.P. (2005): Phosphorus fractions in an acid soil continuously fertilized with mineral and organic fertilizers. Biology and Fertility of Soils, 41: 295–300.
- Wang Y., Chen X., Lu C., Huang B., Shi Y. (2018): Different mechanisms of organic and inorganic phosphorus release from Mollisols induced by low molecular weight organic acids. Canadian Journal of Soil Science, 98: 15–23.
- Yan J., Jiang T., Yao Y., Lu S., Wang Q., Wei S. (2016): Preliminary investigation of phosphorus adsorption onto two types of iron oxide-organic matter complexes. Journal of Environmental Sciences, 42: 152–162.
- Yan X., Wei Z., Hong Q., Lu Z., Wu J. (2017): Phosphorus fractions and sorption characteristics in a subtropical

- paddy soil as influenced by fertilizer sources. Geoderma, 295: 80–85.
- Yang F., He Y.Q., Li C.L., Xu J.B., Lin T. (2006): Effect of fertilization on phosphorus forms and availability in upland red Soil. Acta Pedologica Sinica, 43: 793–799.
- Yang X.Y., Chen X.W., Guo E.H., Yang X.T. (2019a): Path analysis of phosphorus activation capacity as induced by low-molecular-weight organic acids in a black soil of Northeast China. Journal of Soils and Sediments, 19: 840–847.
- Yang X.Y., Chen X.W., Yang X.T. (2019b): Effect of organic matter on phosphorus adsorption and desorption in a black soil from Northeast China. Soil and Tillage Research, 187: 85–91.
- Zhang G.S., Xue J.X., Ni Z.W., Li J.C. (2018): Phosphorus accumulation and sorption characteristics of P-enriched soils in the Dian Lake basin, southwestern China. Journal of Soils and Sediments, 18: 887–896.

Received: November 5, 2019 Accepted: September 10, 2020 Published online: November 23, 2020